

PRI-8800

Automatic Varying Temperature Incubations and Continuous Soil Respiration Measurements System



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A reliable and precise estimate of the temperature sensitivity ($Q_{(0)}$) of soil organic matter (SOM) decomposition is critical to predict feedbacks between the global carbon cycle and climate change. Traditional methods for estimating $Q_{(0)}$ includes CDM mode (constant temperature incubation and discontinuous measurements) and VDM mode (varying temperature incubation and discontinuous measurements). Combining rapidly VCM mode (varying temperature incubations and continuous measurements), PRI-8800 (patented) leads a new method for $Q_{(0)}$ estimation. VCM mode eliminates the underestimated errors by both CDM and VDM modes, provide a more accurate and rapid estimation of the temperature response of SOM decomposition and can be used for large-scale estimation of $Q_{(0)}$.

PRI-8800 is a new and exciting solution for continuous soil respiration measurements combining varying temperature incubations in lab. Excellent compatibility and extensibility with other analyzers such as various GHGs and trace gas concentration analyzers, various isotope analyzers and other gas measurements devices. Applications are related to soil respiration, biodegradability of plastics in solid medium, biodegradability of plastics in aqueous medium, organic waste (solid or liquid samples), food industry, compost biological activity, wastewaters, R&D in biotechnology, biology, ecology and pharmacy.



Key Feature

Varying temperature incubations and continuous measurements Excellent compatibility and extensibility with various analyzers 4, 9 or 16 channels (standard), costomized system is required Automatic temperature control (-20 \sim 80 °C), precision better than 0.1 °C Dual gas circuit to eliminate effect of initial high concentration Inherent cahnnels for isotope and concentration calibration

Specifications

Parameter	Specifications (standard)	Customized	
Control Module			
System Response time	<4s		
Calibration Channels	3		
Power	220 VAC, <350 W		
Dimensions	50 cm × 42 cm × 20 cm		
Sampling Module			
Flask Capacity	150 mL	•	
Precision	±0.05% (Pressure sensor);		
	±0.15℃ (Temp. sensor)		
Flow Rate	1 L/min (0-4 L/min adjustable)		
Rated Track Length	40 cm × 40 cm ×15 cm		
Gas Tube	1/8" Stainless or Teflon	•	
CO ₂ Absorption	Soda lime	•	
Dimensions	80 cm × 80 cm × 70 cm (Autosampler)		



Water Cabinet		
Temperature Control Range	-20~80℃	•
Temperature Control Precision	±0.1°C	
Heating Rate	60 s/°C	•
Cooling Rate	90 s/°C	•
Circulating Pump	20 L/min (adjustable)	•
Autosampler Precision	0.02 mm	
Power Requirements	100-240VAC, 50/60 Hz,	
	1500 W(warming); 1250 W (cooling)	

8800-1 CO ₂ H ₂ O analzyer (Integrated in PRI-8800)		
CO ₂ Accuracy	± 2%	
CO₂ Measurement Range	0-5000 ppm	
H ₂ O Precision (Typical)	± 2%	
H ₂ O Measurement Range	0~100% RH	
Sampling Temperature	-10 ~ 45 °C	
Sampling Pressure	80 ~ 115 kPa	
Sampling Humidity	0-100% R.H, non-concensing	
Fittings	1/4" Swagelok	

AMBA i3211 CO ₂ isotope analyzer		
δ ¹³ C Precision (1σ)	<0.5% (10) @ 0.25s <0.3% (10) @ 1s <0.08% (10) @ 60s <0.05% (10) @ 300s	
CO ₂ Measurement Range	0-10000 ppm	
Measurement Frequency	4Hz or 1Hz	
Sample Flowrate	15 mL/min or 5 mL /min	
Cavity Volume	0.1 mL	
Sampling Temperature	-10 to ~ 45 °C	
Mini Vacuum Pump Flowrate	100 sccm@50 Torr	
Sampling Pressure	50 ~ 133 kPa	
Sampling Humidity	0-100% R.H, non-condensing	
Calibration	Automatic online calibration	
Outputs	Digital (RS-232), Ethernet, USB	
Fittings	1/8" Swagelok	
Dimensions	48 cm (W) × 80 cm (D) × 47.5 cm (H)	
Weight	25 kg	
Power Requirements	100-240VAC, 50/60 Hz, <350 W (start-up) , 200 W (stable)	

Configuration

PRI-8800 includes water bath with refrigerator and heater system, autosampler, 16 channel tray, 50 flasks, control box.

8800-1: stanard CO2 H2O analyzer with 2% CO2 accuracy.

We recommed AMBA i3211 CO2 isotope analyzer for isotopic CO2 analysis, ultra-small cavity, fastest response time and better precison.

Publications

- 1. Liu Y, He NP, Xu L, Tian J, Gao Y, Zheng S, Wang Q, Wen XF, Xu XL, Yakov K. 2019. A new incubation and measurement approach to estimate the temperature response of soil organic matter decomposition. Soil Biology & Biochemistry, 138, 107596.
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- 6. Tian J, He NP#, Hale L, Niu SL, Yu GR, Liu Y, Blagodatskava E, Kuzyakov Y, Zhou JZ. 2018. Soil organic matter availability and climate drive latitudinal patterns in bacterial diversity from tropical to cold-temperate forests. Functional Ecology, 32: 61-70.
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- 13. Shi Y, Sheng LX, Wang ZQ, Zhang XY, He NP, Yu Q. 2016. Responses of soil enzyme activity and microbial community compositions to nitrogen addition in bulk and microaggregate soil in the temperate steppe of Inner Mongolia. Eurasian Soil Science, 49(10): 1149-1160.
- 14. Wang Q, He NP, Liu Y, Li ML, Xu L. 2016. Strong pulse effects of precipitation event on soil microbial respiration in temperate forests. Geoderma, 275: 67-73.
- 15. Wang Q, He NP, Yu GR, Gao Y, Wen XF, Wang RF, Koerner SE, Yu Q. 2016. Soil microbial respiration rate and temperature sensitivity along a north-south forest transect in eastern China: Patterns and influencing factors. Journal of Geophysical Research: Biogeosciences, 121: 399-410.
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Manufacturer: PRI-ECO